Fatigue during ~1-10 min of high-intensity sports/events is multifactorial, but there are strong mechanistic underpinnings demonstrating skeletal muscle acidosis, via accumulating hydrogen ion (H+), to be a key performance limiter. Accordingly, skeletal muscle has various innate intra- and extra-cellular buffering mechanisms to address exercise-induced acidosis.

Carnosine is a key intra-cellular buffer due to its nitrogen containing imidazole side ring, which can accept (buffer) H+ and slow the decline in muscle pH during intense exercise and contribute as much as ~15% to total buffering capacity.

Beyond its role in buffering, carnosine has also been shown to be a diffusible calcium (Ca2+)/H+ exchanger, delivering Ca2+ back to the sarcoplasmic reticulum and H+ away to the cell membrane, suggesting that it might also enhance muscle Ca2+ sensitivity and contraction efficiency.

Carnosine in muscle is synthesized by carnosine synthase, in which the plasma beta-alanine concentration is the rate limiting substrate; consistent data has demonstrated that ~3-6 g beta-alanine daily over at least 4 weeks of supplementation can augment muscle carnosine stores by 30-60%.

Several meta-analyses have shown moderate effect sizes for exercise capacity, and smaller effect sizes for exercise performance over ~1-10 min duration. This translates into ~2-3% performance benefits in non-elite subjects, but ~0.5-1% increases in elite subjects. However, more data is required in elite athlete cohorts.

Despite significant increases in scientific knowledge regarding beta-alanine supplementation protocols and performance efficacy since 2006, there are many prevailing questions and future applied research directions that remain.

INTRODUCTION
Athletes participating in high-intensity sports/events (~1-10 min of all-out effort), or in sports where athletes are required to make repeated high-intensity efforts, have unique performance determinants. All of these sports use large amounts of anaerobically derived adenosine triphosphate (ATP) energy production from phosphocreatine and anaerobic glycolysis, the latter resulting in large amounts of lactate accumulation (>10 mmol/L). The diversity of these unique high-intensity performance determinates is especially prevalent in the middle-distance/high-intensity based events, which feature a blend of aerobic, anaerobic and neuromuscular/mechanical characteristics (Sandford & Stellingwerff, 2019) and many of these determinates are linked to nutritional interventions (Stellingwerff, Bovim, & Whitfield, 2019). Indeed, the limits to high-intensity performance are multifactorial, but a major limiting factor is the ability to tolerate ever increasing levels of muscle acidosis; both intra- and extra-cellular. To enhance extra-cellular buffering, sodium-bicarbonate loading has been researched and utilized by athletes for many decades.

However, it was only in the mid-2000’s that pioneering work by Prof. Roger Harris and colleagues (2006) demonstrated that augmenting intra-cellular (within muscle) buffering was also possible via chronic (several weeks) beta-alanine supplementation, which significantly increased muscle carnosine (b-alanyl-L-histidine) and high-intensity performance (Hill et al., 2007) (Figure 1). Since this time, there has been an explosion of research examining the efficacy of beta-alanine supplementation to optimally augment the muscle carnosine content and enhance subsequent performance. Therefore, this Sports Science Exchange (SSE) article will first examine the limitations to high-intensity performance, with emphasis on anaerobic metabolism and associated buffering mechanisms, and then examine the science of beta-alanine supplementation protocols and various performance outcomes.

**Figure 1:** Schematic outlining the uptake of beta-alanine and synthesis, storage and proposed mechanisms involving augmented carnosine content. β-alanine, beta-alanine; Carn, carnosine; Ca++, calcium; CN1, serum carnosinase; CN2, carnosine synthase; H+, hydrogen ion; PAT1, proton-assisted amino-acid transporter; PEPT1, peptide transporter 1; PHT1, peptide-histidine transporter 1; TauT, taurine transporter. Figure adapted from (Blancquaert et al., 2015; Matthews et al., 2019; Stellingwerff, Decombaz, et al., 2012).
LIMITATIONS TO HIGH-INTENSITY EXERCISE PERFORMANCE AND MUSCLE CARNOSIGINE MECHANISMS OF ACTION

High-intensity sports/events (~1-10 min of all-out effort) are unique since they are at the crossroads of metabolism and receive large contributions of energy from aerobic and anaerobic metabolism (Figure 2). There are three primary energy systems that simultaneously (although at differing rates) provide ATP to power high-intensity exercise, namely the anaerobic systems (also named substrate level phosphorylation): 1) phosphocreatine (PCr) degradation and 2) “anaerobic” glycolysis, and 3) aerobic metabolism (also named oxidative phosphorylation). During the onset of exercise and during exercise situations with increasing intensity, ATP production from aerobic metabolism cannot match the rate of ATP utilization (100% of high-intensity events), and the shortfall in energy supply is made up by anaerobic metabolism. This energy is provided by the PCr and anaerobic glycolysis systems. The latter has a larger capacity than PCr degradation and provides ATP primarily via the breakdown of glycogen in the glycolytic pathway, resulting in excessive pyruvate production. During extreme intensity exercise the high rate of pyruvate production exceeds the rate at which it can be aerobically oxidized by pyruvate dehydrogenase in the mitochondria via the tricarboxylic acid cycle and electron transport chain (Spriet, Howlett, & Heigenhauser, 2000), leading to the extreme levels of lactate production associated with high-intensity sports (competition and training).

Accordingly, high-intensity efforts lasting ~4 min are conducted at ~20-times resting VO\textsubscript{2} values with resultant blood lactate as high as ~25 mmol/L, and the production of hydrogen ions (H\textsuperscript{+}) causing decreases in human skeletal muscle pH from ~7.2 at rest to 6.6 at exhaustion (Hermansen & Osnes, 1972). Although fatigue is multifactorial, there are certainly strong mechanistic underpinnings demonstrating muscle acidosis, via accumulating H\textsuperscript{+}, to be a key limiter to sustained high-intensity exercise over ~1-10 min (Allen, Lamb, & Westerblad, 2008). It has also been known for 80+ years that carnosine is a key intra-cellular buffer due to its nitrogen containing imidazole side ring, which can accept (buffer) H\textsuperscript{+} (Bate-Smith, 1938), and slow the process of lactate production.

Figure 2: A schematic highlighting the beta-alanine zone of potential ergogenic effect (~1-10 min of high-intensity exercise) across all Olympic sports. Each of the three main ATP producing energy systems is highlighted in the background, demonstrating the significant overlap between sports/events that have a great reliance on anaerobic glycolysis and the potential for beta-alanine supplementation to improve performance. Sports and/or events highlighted in red feature existing published data of competition blood lactate levels greater than 10 mmol/L. Total event/match duration is highlighted by the arrow on the right. The dotted line highlights discontinuous (or non-sustained (<0.5 min) high-intensity) durations in each sport (e.g., short sprints with prolonged aerobic recoveries where lactate levels are <10 mmol/L, such as during baseball or soccer). ATP, adenosine triphosphate; CHO, carbohydrate; log, logarithmic; mid-distance, middle-distance; min, minute; PCr, phosphocreatine; sec, seconds.
The decline in muscle pH during intense exercise (Baguet, Koppo, Pottier, & Derave, 2010). The contribution of normal muscle carnosine levels to total intracellular muscle buffering capacity has been suggested to be ~6-7%, but can reach ~15% of total when augmented via beta-alanine supplementation (Harris & Stellingwerff, 2013). Interestingly, it has also been known for 35 years that sprinters and rowers have nearly double the amount of muscle carnosine than marathon runners, and carnosine strongly correlates with Type II muscle fiber content (Parkhouse, McKenzie, Hochachka, & Ovalle, 1985). Taken together, severe metabolic acidosis is a limiter to high-intensity performance of which muscle carnosine can act as one of many intracellular buffers.

Beyond metabolic acidosis limiting ATP production, there are also biomechanical and structural constraints to high-intensity performance that are often not considered when elucidating the limits to high-intensity performance. For example, many middle-distance/power based sports require extreme morphological adaptations (e.g., exceptional quad girths in sprint cycling and rowing) that result in extraordinary speed and explosive strength and/or power. Obviously performance determinants of these sports also need to be considered from a structural and biomechanical origin (e.g., absolute peak force, rate of force development, body mass, etc. (Weyand, Sandell, Prime, & Bundle, 2010)). Near maximal sustained high-intensity running speed appears to be limited by the minimum time needed (ground contact time or time under tension) to apply large forces, which are again related to fast-twitch Type II muscle fiber content (Weyand et al., 2010) and/or the ability of the muscle to repeatedly generate maximal force. Underpinning these various structural/mechanical determinants of performance are fast-twitch Type II muscle fibers (Type Ila and IIX) (Costill et al., 1976), which have nearly double the muscle carnosine content compared to slow-twitch Type I muscle fibers (Baguet et al., 2011; Parkhouse et al., 1985; Stellingwerff, Anwander, et al., 2012).

Carnosine also has numerous proposed physiological roles, although most of the mechanistic support has been generated via cell-culture and rodent research. Nevertheless, many of these in vitro roles may be at play during rest and exercise in humans as well. All of these potential physiological roles have recently been eloquently reviewed by Matthews et al. (2019) with the strongest prevailing ergogenic mechanism for augmented carnosine in athletes being enhanced intra-cellular pH buffering. Secondary mechanisms also include: (1) calcium (Ca^{2+}) handling, including release, reuptake and Ca^{2+} sensitivity; (2) cytoplasmic Ca^{2+}- H^- exchanger described as the “carnosine shuttle” (Figure 1); (3) potential to regulate bioenergetics leading to an increase in glycolytic flux; (4) scavenger of reactive oxygen species altering oxidative stress; and (5) formation of stable conjugates to prevent reactive aldehydes and lipid peroxidation (non-enzymatic detoxification of reactive aldehydes) from occurring.

An interesting emerging mechanism is the carnosine shuttle hypothesis (Figure 1). In support of the carnosine shuttle hypothesis, recent data in cardiac myocytes has demonstrated that carnosine is not just a buffer, but is also involved in Ca^{2+} and H^- handling from the sarcoplasmic reticulum as a type of “carnosine shuttle” (Figure 1; as reviewed by (Blancquaert, Everaert, & Derave, 2015; Matthews, Artioli, Turner, & Sale, 2019), but requires confirmation in human skeletal muscle.

**BETA-ALANINE SUPPLEMENTATION AND CARNOSINE SYNTHESIS**

Carnosine in muscle is synthesized by carnosine synthase, in which the plasma beta-alanine concentration is the rate limiting substrate. The plasma concentration is <2 μM and the Km (concentration needed for a reaction rate that is 50% of maximum) for its uptake into muscle is ~1.0–2.3 mM). In contrast, L-histidine is present in much higher concentrations in plasma (50 μM) and muscle and has a much lower Km (16.8 μM, Figure 1). In 2006, Harris and colleagues (2006) were the first to demonstrate that oral ingestion of 3.2 or 6.4 g/day of beta-alanine (or an isomolar amount of supplemented carnosine (13.0 g/day)) resulted in a 40-60% increase in muscle carnosine in humans. Since this landmark study, every study that has measured muscle carnosine (either via muscle biopsy or magnetic resonance spectroscopy), where a significant amount of beta-alanine was supplemented (e.g., ~3-6 g beta-alanine/day over at least 4 weeks), has shown a significant increase in muscle carnosine content (for reviews see Perim et al., 2019; Stellingwerff, Decombeaz, Harris, & Boesch, 2012). On average, this has led to a significant ~30-50% increase in muscle carnosine content with ~3-6 g beta-alanine supplemented/day over 4-6 weeks. The washout of augmented skeletal muscle carnosine after the termination of beta-alanine supplementation is very slow, with an estimated washout time of ~14 to 15 weeks after a ~50% increase in muscle carnosine (Baguet et al., 2009; Stellingwerff, Anwander, et al., 2012). By far the biggest contributing factor to carnosine synthesis is the dose and duration of beta-alanine supplementation, as studies have shown that doubling the daily dose of beta-alanine results in about double the amount of carnosine synthesized (and halves the time to get to the same increase) (Church et al., 2017; Stellingwerff, Anwander, et al., 2012).

Contemporary data has also shown several potential ways to increase the efficiency of carnosine synthesis with beta-alanine supplementation, as only ~5% of ingested beta-alanine can be accounted for in muscle carnosine, with the rest having an unknown metabolic fate. These include: (1) taking beta-alanine with a meal (providing an insulin effect), as this resulted in greater carnosine synthesis in the soleus, but not gastrocnemius muscle (Stegen et al., 2013); (2) using slow-release beta-alanine as, compared to the pure form, results in greater carnosine synthesis (Varanoske et al., 2019); and (3) undertaking training/exercise while supplementing (Bex, Chung, Baguet, Achten, & Derave, 2015). However, beyond dose and duration of beta-alanine supplementation to impact on carnosine synthesis, these other influencing factors require more scientific validation. Furthermore, a recent seminal study by Saunders et al. (2017a) which implemented 24 weeks of beta-alanine supplementation at 6.4 g/day with 25 subjects, reported a huge range of individual variability for carnosine synthesis. Five subjects had maximal muscle carnosine contents at week 24, but another 5 subjects already had their peak muscle carnosine content
within the first 12 weeks (Saunders, Painelli, et al., 2017). Finally, it does appear that once muscle carnosine is elevated with a loading phase of beta-alanine, continued supplementation with as little as 1.2 g/day of beta-alanine will maintain the elevated carnosine stores (Stegen et al., 2014). Obviously, our understanding of the determinants of the individual response to beta-alanine supplementation are far from being completely understood (Table 1).

**Beta-alanine: prevailing questions and future applied research directions**

What are the mechanistic reasons, beyond baseline carnosine, for the large individual responses to carnosine synthesis from beta-alanine supplementation (Saunders, Painelli, et al., 2017)?

Since carnosine is fiber type dependent (Baguet et al., 2011) are there fiber type specific responses to performance outcomes or ergogenic potential of beta-alanine based on baseline carnosine content (and by extension, fiber type)?

Are there any implications (e.g., enzymatic, protein or transporter attenuation) of long-term beta-alanine supplementation? (>24 weeks, Saunders, Painelli, et al., 2017)?

Does the carnosine shuttle hypothesis (Fig. 1), as mainly developed in cell-culture models, also occur in vivo in humans via augmented H+ and/or Ca2+ handling? How do these separate, but linked, mechanisms impact on performance outcomes in humans?

More training studies are required examining the training adaptation efficacy of beta-alanine supplementation causing carnosine increases – does increased “work” cause increased training adaptations or does a potential attenuation of H+/acidosis impact adaptation?

Given that human skeletal muscle carnosine assessment via non-invasive proton magnetic resonance spectroscopy measurements can be used to grossly fiber type athletes (Baguet et al., 2011) are there distinct fiber type differences in all aspects of training and recovery? How does beta-alanine supplementation impact these parameters?

Are there any pace judgment changes (tolerance) that occur when altering muscle carnosine contents that need careful consideration in time-trial effects such as cycling, swimming and rowing?

**Table 1: Prevailing questions and future applied research directions examining the efficacy of beta-alanine supplementation to improve high-intensity performance and/or training adaptation.**

When beta-alanine is supplemented at levels greater than 800 mg/dose it results in mild paresthesia in most subjects (minor “pins and needles,” skin vasodilation and flushing over ~60-120 min). However, a commercialized slow release beta-alanine tablet has been produced and demonstrated less urinary spillover and a blunting of the peak plasma curve (same area under the curve), with none of the subjects reported any paresthesia symptoms as compared to control (Decombaz, Beaumont, Vuichoud, Bouisset, & Stellingwerff, 2012). Other than a downregulation of the beta-alanine muscle transporter gene and its subsequent protein TauT (Saunders, Painelli, et al., 2017), no other side effects or changes to any blood measurements or acute body weight changes have been demonstrated with beta-alanine supplementation (Harris et al., 2006; Stellingwerff, Anwander, et al., 2012).

**PERFORMANCE OUTCOMES OF AUGMENTED CARNOSINE VIA BETA-ALANINE SUPPLEMENTATION**

The first beta-alanine human performance study was published in 2007 and demonstrated that an increase in muscle carnosine (via beta-alanine supplementation) resulted in dose-response increases in total work done time to exhaustion (TTE) at 110% of wattage maximum (Wmax) at week 4 (+12% TTE) and week 10 (+16% TTE) compared to baseline (Hill et al., 2007). In the following years, a plethora of performance-based studies have demonstrated the efficacy of beta-alanine supplementation to enhance high-intensity outcomes, including:

1. pre-supplementation baseline muscle carnosine being correlated to ~16  s, ~1.5 min, and ~6.5 min rowing performance (Baguet, Bourgois, Vanhee, Achten, & Derave, 2010), showing the importance of muscle carnosine to high-intensity performance;

2. significantly improved 30 s sprint performance (+5% mean power) after prolonged 110 min of cycling (Van Thienen et al., 2009); and

3. enhanced repeated Wingate outcomes (total work done over 4 Wingate tests) in both trained and untrained subjects (de Salles Painelli et al., 2014). However, some studies have demonstrated no performance benefits after beta-alanine supplementation as compared to control, but generally with either supplementation featuring less than ~150 g of total supplemented beta-alanine (Derave et al., 2007; Smith et al., 2009) or with very short, explosive performance protocols under ~30 sec of duration (e.g., 1 repetition maximum, or sprinting), where acidosis will not be limiting (Kendrick et al., 2008; Sweeney, Wright, Glenn Brice, & Doberstein, 2010).

There have been two definitive meta-analyses examining the performance effects of chronic beta-alanine supplementation. The first meta-analysis was published in 2012 and featured 15 studies (n=360 subjects) with an overall outcome of beta-alanine improving the exercise outcome measures compared to control (p=0.002 with an overall effect size (ES) of 0.374) and the majority of the performance effects being demonstrated over 1 to 4 min of high-intensity exercise (Hobson, Saunders, Ball, Harris, & Sale, 2012). However, this meta-analysis was dominated by exercise capacity/TTE protocols (n=13 studies; p=0.013, −2.3% improvement) compared to exercise performance/time-trial (TT) protocols (n=2 studies; p=0.204). It is important to note that effect sizes tend to be greater in duration based TTE protocols, as compared to TTs where intrinsic pacing can also play a role in performance outcomes. A follow-up meta-analysis was completed in 2017 featuring 40 studies (n=1461 subjects), where the overall effect size was 2.1-times smaller (ES=0.18) compared to the 2012 meta-analysis (Saunders, Elliott-Sale, et al., 2017). Interestingly, the ES for exercise capacity/TTE was 0.4998, which was similar to the 2012 meta-analysis, but the ES for performance/TT outcomes was only 0.1078. Again, meta-regression analysis showed that exercise duration of ~1-10 min demonstrated the greatest performance benefits with beta-alanine supplementation (p=0.004; Figure 2).

Finally, there has been some emerging data suggesting smaller overall performance effects (~0.5-1% improvement in performance) of beta-alanine supplementation in well-trained subjects compared to untrained subjects (Bellinger, Howe, Shing, & Fell, 2012; Saunders, Elliott-Sale, et al., 2017). This has been demonstrated in other meta-analyses in the past and probably represents a biological performance ceiling effect found with elite athletes approaching a genetic zenith. Figure 2 highlights the various energy systems, and the performance “sweet spot” of ~1-10 min of duration and the various Olympic sports that,
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References